

There are different P5 primers for PE and single read flowcells, attached at their 5' ends to the flowcell.

P5 PairedEnd: 5'-TTTTTTTTT**AATGATACGGCGACCACCGAGA**UCTACAC-3'

P5 SingleRead: 5'-TTTTTTTTT-(diol)3-**AATGATACGGCGACCACCGA**-3'

#### TruSeq Review:

Single Indexing Truseq "Y" Adapter (Index #1/I7 = ATCACG):

**AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATC**\*T  
GTTCTGTTCTGCGGTATGCTCTAG**CACTA**CACTGACCTCAAGTCTGCAC**ACGAGAAGGCTAG**-P

Double-stranded Standard TruSeq Library Molecule post PCR:

**AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT**-NNN-**AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC****ATCACGATCTCGTATGCCGTCTTCTGCTTG**  
**TTACTATGCCGCTGGTGGCTCTAGATGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGA**-NNN-TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTGTAGTGCT**TAGAGCATAACGGCAGAAGACGAAC**

Denatured ss-Library molecules with read primers:

Read 1 primer ---->	Index read 1 primer ---->
ACACTCTTTCCCTACACGACGCTCTTCCGATCT	GATCGGAAGAGCACACGTCTGAACTCCAGTCAC
TTACTATGCCGCTGGTGGCTCTAGATGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGA-NNN-TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG <b>TAGTGCTAGAGCATAACGGCAGAAGACGAAC</b> --   f/c	

[PE turnaround]

|--**AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT**-NNN-**AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC****ATCACGATCTCGTATGCCGTCTTCTGCTTG** 3'  
TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG  
<---- Read 2 primer

Double-indexed adapter design suitable for sequencing with "Nextera" protocol on MiSeq with a PE flowcell (as of 6/2012)

Annealed adapter example

5'-**AATGATACGGCGACCACCGAGA**TCTACAC**TAGATCGC**ACACTCTTTCCCTACACGAC**GCTCTTCCGATC**\*T N501\_truseq  
3'-GTTCTGTTCTGCGGTATGCTCTA**AGCGAAT**CACTGACCTCAAGTCTGCAC**ACGAGAAGGCTAG**-P N701\_truseq

Double-stranded Library Molecule post PCR (as an example this is a 701-501 combination)

**AATGATACGGCGACCACCGAGATCTACAC****TAGATCGC**ACACTCTTTCCCTACACGAC**GCTCTTCCGATCT**-NNN-**AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC****TAAGGCGA**ATCTCGTATGCCGTCTTCTGCTTG  
**TTACTATGCCGCTGGTGGCTCTAGATGTGATCTAGCGTGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGA**-NNN-TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTGAT**TCCGCT****TAGAGCATAACGGCAGAAGACGAAC**

Denatured ss-Library molecules with read primers:

--Note that index2 is primed from the flow cell primer, POST CLEAVAGE of the 7bp leader sequence. This is why the Nextera recipe requires 7-dark cycles to skip the constant region before reading index2.

<b>AATGATACGGCGACCACCGAGA</b> (index2primer)		
P5 Flowcell Cluster primer-->	Read 1 primer ---->	Index read 1 primer ---->
<b>AATGATACGGCGACCACCGAGA</b> UCTACAC	ACACTCTTTCCCTACACGACGCTCTTCCGATCT	GATCGGAAGAGCACACGTCTGAACTCCAGTCAC
3' TTACTATGCCGCTGGTGGCTCTAGATGTG <b>ATCTAGCC</b> TGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGA-NNN-TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG <b>ATTCCGCT</b> <b>TAGAGCATAACGGCAGAAGACGAAC</b> --   f/c		

[PE turnaround]

|--**AATGATACGGCGACCACCGAGATCTACAC****TAGATCGC**ACACTCTTTCCCTACACGAC**GCTCTTCCGATCT**-NNN-**AGATCGGAAGAGCACACGTCTGAACTCCAGTCAC****TAAGGCGA**ATCTCGTATGCCGTCTTCTGCTTG 3'  
TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG  
<---- 3' Read 2 primer

Example sample sheet:

```
[Header]
IEMFileVersion,3
Investigator Name,ECO
Project Name,DOUBLEINDEX
Experiment Name,EXPT
Date,2012 06/05/12 21:28:11
Workflow,Resequencing
Assay,Nextera
Description
Chemistry,Amplicon
[Reads]
151
151
[Settings]
OnlyGenerateFASTQ,1
[Data]
Sample_ID,Sample_Name,Sample_Plate,Sample_Well,Sample_Project,index,I7_Index_ID,index2,I5_index_ID,Description,GenomeFolder
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2,LS594,RE1018,A01,Exp135,CGTACTAG,N702,TAGATCGC,N501,,C:\Illumina\MiSeq Reporter\Genomes\PhiX\Illumina\RTA\Sequence\Chromosomes
3,LS594,RE1018,A01,Exp135,AGGCAGAA,N703,TAGATCGC,N501,,C:\Illumina\MiSeq Reporter\Genomes\PhiX\Illumina\RTA\Sequence\Chromosomes
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